## RESEARCH

Stem Cell Research & Therapy

**Open Access** 

# Identifying optimal candidates for autologous peripheral blood stem cell therapy in patients with decompensated liver cirrhosis: a prognostic scoring system

Siyuan Tian<sup>1+</sup>, Guanya Guo<sup>1+</sup>, Xia Zhou<sup>1+</sup>, Yansheng Liu<sup>1</sup>, Gui Jia<sup>1</sup>, Linhua Zheng<sup>1</sup>, Lina Cui<sup>1</sup>, Kemei Wang<sup>1</sup>, Miao Zhang<sup>1</sup>, Keshuai Sun<sup>2</sup>, Shuoyi Ma<sup>1</sup>, Chunmei Yang<sup>1</sup>, Xinmin Zhou<sup>1</sup>, Changcun Guo<sup>1\*</sup>, Yulong Shang<sup>1\*</sup> and Ying Han<sup>1\*</sup>

## Abstract

**Background** Stem cell transplantation shows great potential to improve the long-term survival of cirrhosis patients. However, therapeutic effects may not be homogeneous across the whole study population. This study constructed an easy-to-use nomogram to improve prognostic prediction and aid in treatment decision making for cirrhotic patients.

**Methods** From August 2005 to April 2019, 315 patients with decompensated cirrhosis receiving autologous peripheral blood stem cell (PBSC) transplantation were enrolled in this study. They were randomly classified into training (2/3) and validation (1/3) groups. A predictive model was developed using Cox proportional hazard models and subsequently validated. The predictive performance of the model was evaluated and also compared with other prognostic models.

**Results** Age, creatinine, neutrophil-to-lymphocyte ratio, and Child–Turcotte–Pugh class were included in the nomogram as prognostic variables. The nomogram showed high discrimination power concerning the area under receiver operating characteristic curves (3/5-year AUC: 0.742/0.698) and good consistency suggested by calibration plots. Patients could be accurately stratified into poor- and good-outcome groups regarding liver-transplantation free survival after receiving PBSC therapy (P < 0.001). Compared with poor-outcome group, the liver function of patients listed for liver transplantation in the good-outcome group was significantly improved (P < 0.001). Besides, our nomogram achieved a higher C-index (0.685, 95% CI 0.633–0.738) and better clinical utility compared with other conventional prognostic models.

<sup>†</sup>Siyuan Tian, Guanya Guo and Xia Zhou contributed equally to this work.

\*Correspondence: Changcun Guo guochc@sina.com Yulong Shang shangyul870222@163.com Ying Han hanying1@fmmu.edu.cn Full list of author information is available at the end of the article



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.gr/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/.

**Conclusions** The proposed nomogram facilitated an accurate prognostic prediction for patients with decompensated cirrhosis receiving PBSC transplantation. Moreover, it also held the promise to stratify patients in clinical trials or practice to implement optimal treatment regimens for individuals.

Keywords Stem cell therapy, Liver cirrhosis, Nomogram, Risk stratification

## Background

According to the Global Burden of Disease Report, nearly 1.32 million fatalities were attributed to liver cirrhosis in 2017, representing 2.4% of global all-cause mortality [1]. Moreover, the prevalence of decompensated cirrhosis presents a persistently increasing trend during the past 30 years, and the 5-year mortality rate due to severe complications of advanced cirrhosis can be as high as 18–80% [2]. At present, liver transplantation remains the only definitive treatment for patients with decompensated cirrhosis [3]. However, only a minority of the patients could receive transplantation due to donor shortage, and the clinical outcome is still not satisfactory [4].

New possibilities for treating cirrhosis have emerged due to recent advancements in stem cell biology [5, 6]. Stem cells are characterized by the capacity of selfrenewal, immunomodulation, and multipotent differentiation [7]. In 2005, stem cell transplantation was first utilized for liver disease, which provided a novel therapeutic approach in the clinical setting [8]. Stem cell-based therapies have since advanced rapidly and attracted significant interest in the field of hepatology, including liver cirrhosis. Among them, peripheral blood stem cell (PBSC) transplantation stands out as a feasible option because of its low risk and high tolerability for patients [9]. Most of the previous studies have confirmed that stem cell therapy could improve liver biochemical parameters [10–13]. However, some researchers drew a contrary conclusion, like the study conducted by Newsome et al. [14]. On the one hand, the small sample size and lack of assessment of long-term clinical outcomes prevent definitive conclusions regarding the effect of this treatment on cirrhosis. On the other hand, the heterogeneities of the patients varying in etiology, course of the disease as well as immune status should also be taken into full consideration, which may give rise to these varied clinical observations and treatment outcomes of individual patients. In contrast, our previous studies with up to 10 years of follow-up confirmed that patients with decompensated cirrhosis had an elevated chance of survival after PBSC transplantation [15]. However, there is no individual prognostic tool to provide survival estimates for patients with cirrhosis after stem cell therapy to guide individualized therapeutic strategies.

So, we conducted this retrospective cohort study to specifically identify the prognostic factors of patients with decompensated cirrhosis receiving PBSC transplantation, by analyzing demographic data, etiology of liver disease, as well as clinical features. By integrating these variables, we developed and validated a predictive tool, which could facilitate prognostic prediction and inform therapy choice in clinical application.

## Methods

## Study design and population

From August 2005 to April 2019, we recruited 333 patients with decompensated cirrhosis, who completed autologous PBSC transplantation in the Xijing Hospital. Specifically, the criteria for selection were as follows: (1) age range from 18 to 75 years; (2) decompensated cirrhosis (hepatitis B or C); (3) Child–Turcotte–Pugh (CTP) score of 7 or greater; (4) antiviral therapy for a minimum of 6 months prior to PBSC transplantation. Patients were excluded based on the following: (1) previous diagnosis within the last 30 days of spontaneous peritonitis, hepatic encephalopathy, or variceal hemorrhage; (2) hepatorenal syndrome; (3) hepatocellular carcinoma or other malignancies; (4) alcohol abuse. Clinical symptoms, upper digestive endoscopy, ultrasonography, or computerized tomography were used to diagnose decompensated cirrhosis [16]. Chronic hepatitis B or C was diagnosed based on patient history, serologic tests, and biochemical indicators. Patients with unknown survival information were excluded from the analysis. All of the eligible patients were randomly assigned a number by a computer program, and thus they were classified into the training or the validation group at a 2:1 ratio. Informed consent was obtained from all patients and the study was approved by the Ethics Committee of Xijing Hospital.

## **Treatment procedures**

The procedure of PBSC transplantation was described previously [15]. In brief, G-CSF was administered to the patients subcutaneously once per day at a dosage ranging from 5 to 10 mg/kg/d for a total of 4 consecutive days. Leukapheresis with the COBER Spectra Apheresi System (Gambro BCT, Inc, Stockholm, Sweden) was employed to harvest the mobilized PBSCs. Then, patients received 50 mL of leukapheresis products comprising  $10^7-10^8$  cells per kg of body weight administered into their hepatic arteries. Following leukapheresis, the cell preparations were examined by flow cytometry to

assess the percentages of CD14<sup>+</sup> and CD34<sup>+</sup> cells. CD14<sup>+</sup> cells accounted for around 90% of the newly extracted PBMCs, whereas CD34<sup>+</sup> cells accounted for 20% of the total cells. There were at least 50 ml of newly extracted PBMCs comprising  $6 \times 10^8$  cells. In each procedure, over  $1 \times 10^8$  CD34<sup>+</sup> cells were transplanted into the recipient's liver via the hepatic artery. Every participant received only one PBSC treatment. The procedure protocol was standardized and strictly followed. All patients received standard medications like antiviral treatment, and complications such as hepatoencephalopathy, hepatorenal syndromes, variceal hemorrhage, and ascites were managed in compliance with updated guidelines on the basis of PBSC transplantation [16–18].

#### Data collection and outcomes

The electronic medical records were retrieved for all pertinent clinical and demographic information, such as age, sex, etiology of liver disease, blood routine test, liver, and kidney function test, as well as coagulation parameters. As described previously [15], biochemical indexes were set as dichotomous variables, with the reference being the normal range of healthy people. The neutrophil-tolymphocyte ratio (NLR) was defined as the absolute neutrophil count divided by the absolute lymphocyte count. The X-tile software (version 3.6.1) was used to identify the optimal cutoff values for NLR, which were then transformed into categorical variables. The CTP [19], ALBI [20], and model for end-stage liver disease (MELD) [21] score were calculated according to the relevant formulas. Baseline data were collected during patients' first visit and they were regularly followed up by outpatient visits or telephone calls. The end point of each patient was recorded as liver transplantation (LT) or death. Patients who remained alive without LT at last follow-up date (January 1st, 2020) or lost to follow up were censored. LT-free survival was defined as the duration between the initiation of PBSC treatment and liver transplantation or death/censure.

## Statistical analysis

SPSS 23.0 software (SPSS, IBM) and R software version 4.1.1 were applied in analyzing statistical data. With the use of X-tile software (version 3.6.1), we derived the optimal cutoff value for NLR and risk score. Median with interquartile range (IQR) and/or mean  $\pm$  (SD) were employed to present continuous variables. As deemed appropriate, we contrasted continuous variables by means of the Student's t test or the Mann–Whitney U test. Quantitative comparisons of categorical variables, which were expressed as numbers with percentages, were made using the  $\chi$ 2 test or Fisher's exact test. For paired data that did not follow a normal distribution,

a Wilcoxon-matched pair test was applied. To determine independent prognostic factors linked to LT-free survival, the Cox proportional hazards model was used. In the subsequent multivariate analysis, we took into account the factors that were significant in the previous univariate analysis. The predictive model was finally determined by the backward and forward stepwise approach based on the Akaike information criterion (AIC). A Kaplan-Meier (KM) curve was generated, and the log-rank test was utilized to determine the differences (variations) in survival rates. Then, using the "rms" package included in R software, a nomogram was developed as per the results of a multivariate analysis carried out on the training dataset. Measurements of discrimination and calibration were taken from the training dataset as well as the validation dataset to further analyze the performance of the nomogram. The time-dependent ROC curves were plotted with the "timeROC" R package, and 3- and 5-year AUC values of the ROC curve were calculated. In addition, the calibration curve was employed for visualization, and within the curve, the 45-degree line was found to provide the most accurate prediction. Moreover, the discriminatory ability of the nomogram was compared with three conventional prognostic models for liver disease (CTP, ALBI, and MELD score) by calculating C-index. The clinical significance of these models was evaluated with the help of decision curves by quantifying the net benefits at different probability thresholds. A P-value (two-sided) of < 0.05 denoted the significance threshold.

## Results

## **Baseline characteristics**

During the study period, a total of 333 patients met the inclusion and exclusion criteria, 18 patients were excluded due to incomplete survival information. Finally, 315 eligible patients (training cohort: N1 = 210, validation cohort: N2 = 105) were included in the analysis (Additional file 1: Fig. S1). The patterns of other missing values are illustrated in Additional file 1: Fig. S2. The baseline characteristics of all patients are summarized in Table 1. The median follow-up was 50.5 (IQR 13.6-86.5) months and 47.6 (IQR 17.3–81.5) months in the training and validation cohorts, correspondingly. Patients were predominantly male (69.8%) with a median age of 48 (IQR 42–56) years. The hepatitis B virus accounted for the majority of the etiology of liver cirrhosis. No significant variation was found between the training cohort and validation cohorts in terms of demographic data and other clinicopathological parameters except that patients in the training cohort recorded a slightly elevated level of serum bilirubin. The LT-free survival of patients was displayed in Additional file 1: Fig. S3, and no significant variation existed across the groups. In the entire cohort, the median value of NLR

| Baseline characteristics         | Entire cohort                          | Training cohort    | Validation cohort | P-value |
|----------------------------------|--|--------------------|-------------------|---------|
|                                  | (N=315)                                | (N1=210)           | (N2 = 105)        |         |
| Condor n (%)                     |  |                    |                   | 0.224   |
| Female                           | 95 (30 2)                              | 68 (32.4)          | 27 (25 7)         | 0.224   |
| Male                             | 220 (69.8)                             | 142 (67 6)         | 78 (74 3)         |         |
|                                  | 48 (42-56)                             | 48 (42-56)         | 47 (41-56)        | 0.512   |
| Etiology n (%)                   | -10 (-12 - 50)                         | 10 (12 50)         | - (+ 50)          | 0.669   |
| HBV                              | 288 (91 4)                             | 191 (91 0)         | 97 (92.4)         | 0.005   |
| HCV                              | 27 (86)                                | 19 (9 0)           | 8 (7 6)           |         |
| PIT 10 <sup>9</sup> /I           | 27 (0.0)                               | 19 (9.0)           | 0 (7.0)           | 0.805   |
| > 100                            | /3 (13 7)                              | 20 (13.8)          | 14 (13 3)         | 0.095   |
| < 100                            | 271 (86.0)                             | 29 (13.0)          | 91 (86 7)         |         |
|                                  | 271 (00.0)                             | 100 (05.7)         | 91 (00.7)         | 0.375   |
| < 40                             | 101 (E7 E)                             | 117 (EE 7)         | 64 (61 0)         | 0.575   |
| ≤ 40<br>> 40                     | 124 (42 5)                             | 02 (44.2)          | 41 (30.0)         |         |
| >40                              | 134 (42.5)                             | 95 (44.5)          | 41 (39.0)         | 0.120   |
| AST, IU/L, /I (%)                | 161 (511)                              | 101 (40.1)         | 60 (57.1)         | 0.150   |
| ≤ 54                             | 101 (51.1)                             | 101 (48.1)         | 60 (57.1)         |         |
| > 54                             | 154 (48.9)                             | 109 (51.9)         | 45 (42.9)         | 0.004   |
| Albumin, g/L, <i>n</i> (%)       | 114 (26.2)                             |                    | 20 (27.1)         | 0.804   |
| ≥30                              | 114 (36.2)                             | /5 (35./)          | 39 (37.1)         |         |
| < 30                             | 201 (63.8)                             | 135 (64.3)         | 66 (62.9)         | 0.000   |
| I BIL, μmol/L, <i>n</i> (%)      | ()                                     |                    |                   | 0.032   |
| ≤1/.1                            | 52 (16.5)                              | 28 (13.3)          | 24 (22.9)         |         |
| > 17.1                           | 263 (83.5)                             | 182 (86.7)         | 81 (77.1)         |         |
| Creatinine, µmol/L, <i>n</i> (%) |  |                    |                   | 0.755   |
| ≤106                             | 293 (93.0)                             | 196 (93.3)         | 97 (92.4)         |         |
| >106                             | 22 (7.0)                               | 14 (6.7)           | 8 (7.6)           |         |
| Sodium, mmol/L, <i>n</i> (%)     |  |                    |                   | 0.946   |
| ≥137                             | 242 (76.8)                             | 160 (76.2)         | 82 (78.1)         |         |
| <137                             | 67 (21.3)                              | 44 (21.0)          | 23 (21.9)         |         |
| PT (s)                           |  |                    |                   | 0.444   |
| ≤15                              | 23 (7.3)                               | 17 (8.1)           | 6 (5.7)           |         |
| >15                              | 292 (92.7)                             | 193 (91.9)         | 99 (94.3)         |         |
| Fibrinogen, g/L, <i>n</i> (%)    |  |                    |                   | 0.875   |
| ≥2                               | 232 (73.7)                             | 155 (73.8)         | 77 (73.3)         |         |
| <2                               | 82 (26.0)                              | 54 (25.7)          | 28 (26.7)         |         |
| INR                              |  |                    |                   | 0.293   |
| ≤ 1.5                            | 131 (41.6)                             | 83 (39.5)          | 48 (45.7)         |         |
| > 1.5                            | 184 (58.4)                             | 127 (60.5)         | 57 (54.3)         |         |
| NLR                              | 1.53 (1.02–2.48)                       | 1.50 (1.01–2.44)   | 1.61 (1.06–2.68)  | 0.233   |
| Ascites, n (%)                   |  |                    |                   | 0.832   |
| None                             | 44 (14.0)                              | 31 (14.8)          | 13 (12.4)         |         |
| Mild                             | 123 (39.0)                             | 82 (39.0)          | 41 (39.0)         |         |
| Moderate to massive              | 148 (47.0)                             | 97 (46.2)          | 51 (48.6)         |         |
| CTP class, n (%)                 |  |                    |                   | 0.686   |
| В                                | 185 (58.7)                             | 125 (59.5)         | 60 (57.1)         |         |
| С                                | 130 (41.3)                             | 85 (40.5)          | 45 (42.9)         |         |
| CTP score                        | 9.0 (8.0-11.0)                         | 9 (8.0-11.0)       | 9 (8.0–11.0)      | 0.955   |
| MELD score                       | 14.2 (11.4–17.2)                       | 14.2 (11.9–17.0)   | 14.3 (10.5–17.7)  | 0.561   |
| Al Bl score                      | -1.42+0.40                             | -1.43+0.38         | -1.43+0.44        | 0.160   |
| Survival status, n (%)           |  |                    |                   | 0.294   |
| Alive                            | 182 (578)                              | 117 (55 7)         | 65 (61 9)         | 0.22    |
| Dead/liver transplantation       | 129 (41 0)/4 (1 2)                     | 90 (42 9)/3 (1 4)  | 39 (37 0)/1 (1 0) |         |
| Follow-up duration months        | 129 (T1.0)/T (1.2)<br>20 5 (15 8_96 A) | 50 (72.2)/ 5 (1.7) | Δ76 (17 3_ Q1 5)  | 0.000   |
|                                  | (0.00-0.0)                             | (0.00-0.01)        | (6.10-6.11) 0.1+  | 0.902   |

## Table 1 Demographic data and baseline characteristics of the study cohort

## Table 1 (continued)

HBV hepatitis B virus, PLT platelet, ALT alanine aminotransferase, AST aspartate aminotransferase, TBIL total bilirubin, PT prothrombin time, INR international normalized ratio, NLR neutrophil-to-lymphocyte ratio, CTP Child–Turcotte–Pugh, MELD model for end-stage liver disease, ALBI albumin–bilirubin

was 1.53 (IQR 1.02–2.48). The optimal cutoff value of NLR is 2.7, as determined by X-tile software. Compared with patients with NLR > 2.7, patients with NLR  $\leq$  2.7 had a more favorable prognosis (Additional file 1: Fig. S4).

## Prognostic factors for LT-free survival

Univariate and multivariate regression analysis were conducted in the training cohort to identify prognostic factors for patients receiving PBSCs transplantation. The results of univariate analysis revealed that age, creatinine, NLR, CTP class, and MELD score were significantly associated with the prognosis of patients. Subsequently, these variables were imported into a multivariate stepwise Cox regression analysis to ascertain the extent of their independent influence on clinical outcomes (Table 2). The AIC was computed to evaluate the relative goodness of fit and the simplicity of the model before making the final selection. Finally, we picked the dominant model containing all four variables, since it had the lowest AIC value (Additional file 1: Table S1). Furthermore, survival curves stratified by these factors are shown in Additional file 1: Fig. S5, demonstrating significant difference in survival.

## Nomogram construction and validation

On the basis of these independent prognostic parameters, a nomogram characterized by scale line and scores weight reflected 3- and 5-year LT-free survival prediction was established (Fig. 1). The score corresponding to each nomogram variable was generated by the weight of each clinical factor in the multivariate regression analysis. The detail score for each variable is listed in Additional file 1: Table S2. We employed ROC curves and calibration curves to evaluate the performance of the nomogram. In the training cohort, the area under the ROC curve for predicting 3- and 5-year LT-free survival of patients was 0.742 and 0.698, respectively (Fig. 2A). Meanwhile, the accuracy of the nomogram remained satisfactory at the time points mentioned above in the validation cohort (Fig. 2C). Moreover, the calibration curves also demonstrated good probability consistencies between the predicted and observed values (Fig. 2B, D).

## Risk stratification and subgroup analysis

By calculating the risk score, patients from the training cohort were stratified into two groups according to the optimal cutoff values determined by the X-tile software (Additional file 1: Fig. S6). Consequently, patients were divided into good-outcome group (score < 144) and poor-outcome group (score  $\ge$  144). To further verify the effectiveness of our model, we compared the survival difference within two risk subgroups in the training, validation and entire cohort. Additionally, Fig. 3A–C depicts that the survival curves were well separated, which indicated good discrimination in different subgroups. The survival status and risk score distribution are displayed in Fig. 3D–F. To reduce the possible risk of bias due to incomplete outcome data, we also performed a subgroup analysis with a complete follow-up of patients. Similarly, a significant difference in survival curves was also observed (Additional file 1: Fig. S7).

To better assess the prognostic ability of the risk score, we performed a stratification analysis to confirm whether it retains its ability to predict LT-free survival in various subgroups. The good-outcome group showed better LTfree survival in both HBV and HCV subgroups in contrast with the poor-outcome group (Additional file 1: Fig. S8A–B). We also verified that the risk score retained its capability to distinguish patients with different clinical outcomes in either HBV-DNA (+) or HBV-DNA (-) patients (Additional file 1: Fig. S8C-D). Moreover, similar results were also obtained in subgroups analysis stratified by liver function classes including ALBI grade, MELD score as well as CTP class (Fig. 4A-F). Notably, among patients with baseline MELD score  $\geq$  15, significant improvements in MELD score were observed in the good-outcome group. However, no significant variation was observed in the poor-outcome group (Fig. 5A). In comparison, good-outcome group showed a higher proportion of MELD score < 15 following PBSC transplantation. The median follow-up time in goodand poor-outcome was 90 (IQR 46-237) and 68 (IQR 40-208) days, respectively. No significant variation was noted between the groups at the follow-up (P=0.282).

## Comparison of the predictive performance and net clinical benefits with other models

The performance of the nomogram and other models including ALBI grade, MELD score, and CTP class were compared by calculating C-index or plotting DCA curves. The C-index value for the entire cohort of the nomogram was 0.685 (0.633–0.738), which was considerably superior in contrast with the other three models (Additional file 1: Table S3). Furthermore, to evaluate the nomogram's clinical significance, we conducted a decision curve analysis to quantify the net benefits. Compared with other models, the nomogram exhibited a satisfactory benefit at different threshold probabilities

| Table 2    | Ur  | nivariate | anc | l multi   | variate | Сох    | regres | sion | ana   | lysi | s of |
|------------|-----|-----------|-----|-----------|---------|--------|--------|------|-------|------|------|
| prognost   | ic  | factors   | for | liver-tra | ansplar | ntatio | n free | sur∖ | /ival | in   | the  |
| training o | coł | nort      |     |           |         |        |        |      |       |      |      |

| Variable                | Univariate cox regr         | ession  | Multivariate cox<br>regression |         |  |
|-------------------------|-----------------------------|---------|--------------------------------|---------|--|
|                         | HR (95% CI) <i>P</i> -value |         | HR (95% CI) <i>P</i> -value    |         |  |
| Gender                  |                             |         |                                |         |  |
| Female                  | Reference                   |         |                                |         |  |
| Male                    | 1.222 (0.788–1.897)         | 0.371   | -                              | -       |  |
| Age (years)             |                             |         |                                |         |  |
| ≤ 50                    | Reference                   |         | Reference                      |         |  |
| > 50                    | 1.838 (1.221–2.767)         | 0.004   | 1.826 (1.207–<br>2 761)        | 0.004   |  |
| Etiology of c           | irrhosis                    |         | 2.0 0 1)                       |         |  |
| HBV                     | Reference                   |         |                                |         |  |
| HCV                     | 0.805 (0.389–1.666)         | 0.560   | -                              | -       |  |
| PLT, 10 <sup>9</sup> /L |                             |         |                                |         |  |
| ≥100                    | Reference                   |         |                                |         |  |
| < 100                   | 1.338 (0.672–2.663)         | 0.407   | -                              | -       |  |
| ALT, IU/L               |                             |         |                                |         |  |
| ≤40                     | Reference                   |         |                                |         |  |
| >40                     | 0.871 (0.578–1.312)         | 0.509   | -                              | -       |  |
| AST, IU/L               |                             |         |                                |         |  |
| ≤54                     | Reference                   |         |                                |         |  |
| >54                     | 1.122 (0.745–1.690)         | 0.581   | _                              | _       |  |
| Albumin, g/L            | - · · ·                     |         |                                |         |  |
| ≥30                     | Reference                   |         |                                |         |  |
| < 30                    | 1.017 (0.667–1.552)         | 0.937   | -                              | -       |  |
| TBIL, µmol/L            |                             |         |                                |         |  |
| ≤17.1                   | Reference                   |         |                                |         |  |
| > 17.1                  | 1.102 (0.612–1.985)         | 0.747   | _                              | _       |  |
| Creatinine, µ           | mol/L                       |         |                                |         |  |
| ≤106                    | Reference                   |         | Reference                      |         |  |
| >106                    | 2.750 (1.316–5.748)         | 0.007   | 2.906 (1.375–<br>6.141)        | 0.005   |  |
| Sodium, mm              | iol/L                       |         |                                |         |  |
| ≥137                    | Reference                   |         |                                |         |  |
| < 137                   | 0.954 (0.574-1.586)         | 0.857   | -                              | _       |  |
| PT (s)                  |                             |         |                                |         |  |
| <15                     | Reference                   |         |                                |         |  |
| >15                     | 1.037 (0.453-2.374)         | 0.932   | -                              | _       |  |
| Fibrinogen, d           | ŋ∕l                         |         |                                |         |  |
| ≥2                      | Reference                   |         |                                |         |  |
| <2                      | 0.956 (0.597–1.531)         | 0.851   | -                              | _       |  |
| INR                     |                             |         |                                |         |  |
| ≤1.5                    | Reference                   |         |                                |         |  |
| >15                     | 0.973 (0.643–1 472)         | 0 895   | _                              | _       |  |
| NIR                     |                             | 0.000   |                                |         |  |
| <27                     | Reference                   |         | Reference                      |         |  |
| >27                     | 3,490 (2,245–5,427)         | < 0.001 | 3.086 (1.973-                  | < 0.001 |  |
|                         | 5.120 (2.2.13 - 5.127)      | . 0.001 | 4.828)                         |         |  |

## Table 2 (continued)

| Variable                         | Univariate cox regres       | Multivariate cox<br>regression<br>HR (95% CI) <i>P</i> -value |                         |       |
|----------------------------------|-----------------------------|---|-------------------------|-------|
|                                  | HR (95% CI) <i>P</i> -value |   |                         |       |
| Ascites, n (%)                   |                             |   |                         |       |
| None                             | Reference                   |   |                         |       |
| Mild                             | 1.064 (0.560–2.022)         | 0.850   | -                       | -     |
| Moder-<br>ate<br>to mas-<br>sive | 1.197 (0.648–2.212)         | 0.567   | -                       | _     |
| CTP class                        |                             |   |                         |       |
| В                                | Reference                   |   | Reference               |       |
| С                                | 1.722 (1.144–2.591)         | 0.009   | 1.734 (1.145–<br>2.626) | 0.009 |
| MELD score                       |                             |   |                         |       |
| < 15                             | Reference                   |   |                         |       |
| ≥15                              | 1.615 (1.075–2.429)         | 0.021   | -                       | -     |
| ALBI grade                       |                             |   |                         |       |
| 2                                | Reference                   |   |                         |       |
| 3                                | 0.973 (0.646–1.464)         | 0.895   | -                       | -     |

HBV hepatitis B virus, PLT platelet, ALT alanine aminotransferase, AST aspartate aminotransferase, TBIL total bilirubin, PT prothrombin time, INR international normalized ratio, NLR neutrophil-to-lymphocyte ratio, CTP Child–Turcotte–Pugh, MELD model for end-stage liver disease, ALBI albumin–bilirubin, HR hazard ratio, CI confidence interval

in predicting 3- and 5-year LT-free survival (Additional file 1: Fig. S9).

## Development of an online calculator for the prognostic evaluation

For user convenience, we also developed a web-based online calculator to assist clinicians in making survival estimates for those patients receiving PBSCs treatment. The online tool is available at: https://xijing-PBSC-model. shinyapps.io/DynNomapp/.

By selecting the corresponding clinical parameters, the predicted survival probability for the whole time period could be obtained. The screenshot of the web-based nomogram for the prediction of LT-free survival is shown in Additional file 1: Fig. S10.

## Discussion

In recent years, results from clinical trials and animal models show that stem cell therapy holds exciting therapeutic promise for hepatic fibrosis [22, 23]. Our previous study also demonstrated that autologous PBSC transplantation could improve the liver functions of patients with cirrhosis [11]. On the basis, we further evaluated the long-term effects of PBSC therapy. The results showed that PBSC transplantation is a safe treatment regimen for patients with cirrhosis and can improve long-term



Fig. 1 Nomogram for predicting the 3- and 5-year LT-free survival of cirrhosis patients receiving PBSC transplantation. Cre Creatinine, NLR neutrophil-to-lymphocyte ratio, CTP Child–Turcotte–Pugh, LT-free survival Liver-transplantation free survival, PBSC peripheral blood stem cell

survival without increasing the risk of hepatocellular carcinoma [15]. A recent randomized controlled trial (RCT) also reported that mesenchymal stem cell therapy substantially enhanced the liver function and long-term survival rate of patients with HBV-associated decompensated liver cirrhosis [24]. However, existing research has not definitively shown the optimal cirrhosis population that would gain the most benefit from stem cell transplantation. In the current investigation, we developed and validated a prognostic model by integrating the superiorities of a large sample cohort with a follow-up duration of more than 10 years, which could accurately predict individual outcomes with favorable performance and discrimination. This scoring system was proposed to stratify patients into good- and poor-outcome groups with significantly different treatment outcomes. It will be helpful to guide clinicians to select the appropriate treatment option and control the heterogeneity of enrolled patients in future clinical trials.

In this study, age older than 50 years, serum creatinine > 106  $\mu$ mol/L, NLR > 2.7, and CTP class C independently functioned as factors linked to the dismal survival of patients after PBSC transplantation. These factors are all easily accessible in the clinical practice. Currently, no related study has compared the therapeutic efficacy of stem cell transplantation on cirrhosis caused by different etiologies. In our study, only viral hepatitis-related cirrhotic patients were enrolled. Considering China's status as a region with a high HBV disease burden, gaining deeper knowledge about the efficacy of stem cell therapy in the treatment of HBV-induced cirrhosis is crucial in improving the prognosis of patients in China. Besides, the performance status and organ function are also important factors affecting the effectiveness of stem cell therapy. Patients with older age, impaired organ function, and infections will just receive a limited survival benefit from the treatment. Similarly, Zhang et al. reported that the therapeutic effects of stem cell therapy for liver cirrhosis varied partly by patient age. Assessing patient age is necessary prior to clinical use of stem cell therapy [25].

NLR is a simply calculated and routinely available marker reflecting systemic inflammation. In our study, the cutoff value of the NLR was set as 2.7. Patients with NLR>2.7 exhibited poor long-term prognosis. It was reported that NLR was intimately linked to the prognosis of patients including those with compensated or decompensated cirrhosis [26]. In addition, this index could also serve as an independent indicator of mortality in patients with liver failure awaiting transplantation [27, 28]. Neutrophils and lymphocytes are key components of the immune system. Elevated neutrophil count reflects ongoing inflammation, whereas decreased lymphocyte count is associated with immunosuppression. In recent years, mounting evidence suggests that the effectiveness of stem cell transplantation is primarily dependent on the microenvironment in vivo during liver damage and may be the key factor in determining how well a therapy will work [29]. We therefore speculate the immunosuppressive condition may be one of the attributes that hinders the efficacy of stem cell therapy.

Based on the above factors, a stepwise model selection procedure based on AIC was applied to construct



Fig. 2 Evaluation of the predictive performance of the nomogram. A and C ROC curves for the prediction of 3- and 5-year LT-free survival, B and D Calibration curves for prediction of 3- and 5-year LT-free survival in the training and validation cohort. AUC area under the curve, ROC receiver operator characteristic, LT-free survival Liver-transplantation free survival

the final nomogram. The nomogram showed good discriminative and calibration ability in both the training and validation cohort. Meanwhile, this individualized risk stratification of nomogram could contribute to identify subgroups where it would be advantageous for using PBSC transplantation to improve survival. The predictive power and clinical utility of the nomogram were also compared with some commonly used prognostic models. Our results further confirmed that this model was superior to CTP class, MELD score, and ALBI grade. The target population is the core issue of prognostic models, since it determines the different treatment or care option. Although the clinical application of stem cells has formally expanded, the recommended candidates have not vet been clearly defined. The heterogeneity of the study population may lead to the inconsistent outcomes of stem cell treatment on cirrhosis. Newsome et al. conducted a randomized controlled study and found that stem cell infusion did not improve liver dysfunction or fibrosis [20]. Notably, the majority of the people who participated in the research were older individuals who had cirrhosis, and the causes of their condition were multiple, including primary biliary cholangitis, non-alcoholic fatty liver disease, and alcohol-related liver disease. Based on our risk stratification model, those patients are likely to be classified as a poor-outcome group if they also suffered from other risk factors. Besides, some scholars pointed out that the dose and viability of transfused stem cells were lower than other studies showing an improvement in liver function and disease severity [30]. So, the result of that trial failed to support the potential efficacy of stem cell therapy. This also suggests to us that not all patients are the best target population and the implementation of stem cell therapy should be scrutinized. The current model may provide risk stratification criteria to identify the optimal candidate of PBSC transplantation.



Fig. 3 Survival analysis of LT-free survival in different risk subgroups stratified by the nomogram score. Kaplan–Meier curves of stratified survival in the A training, B validation and C entire cohorts. The distribution of risk score calculated by the nomogram and survival status of patients in the D training, E validation and F entire cohorts. Good outcome (nomogram score < 144) and poor outcome (nomogram score  $\ge$  144). LT-free survival, liver-transplantation free survival

There is no denying that liver transplantation is still the most effective treatment for patients with endstage liver disease. Nevertheless, most patients do not undergo liver transplantation due to lack of donors, high costs, and immune rejection. As per the available data, nearly 40% of those eligible for a liver transplant have to wait over 2 years, and the lack of livers available for transplantation is directly responsible for the deaths of 10–15% of patients [31, 32]. Moreover, liver transplantation is not feasible for all cirrhosis patients. Since 2002, MELD scoring system has been employed for donor liver allocation in the USA [33]. In particular, patients with MELD scores <15 are not the ideal candidates for liver transplantation [34]. The application of PBSC therapy is expected to fill the gap of treatment. At this time, our risk stratification model could be a useful tool in clinical trials to identify the target populations that could benefit from PBSC transplantation. Then, we could enroll these patients in clinical stem cell studies to reduce their potential risk of disease progression due to long waiting periods for suitable donor livers. After assessing the risk of the persistent shortage of appropriate liver donors, the model might potentially be used to select the optimal patients for PBSC therapy when the MELD score is  $\geq$  15. As seen in our results, 75% of patients among those with a MELD score  $\geq$  15 could be classified into a goodoutcome group. Meanwhile, a remarkable enhancement in the MELD score was noted in the good-outcome group, with 73.4% of treated patients delisted for liver transplantation.

To the best of knowledge, this is the first nomogram for predicting the long-term outcomes of patients with cirrhosis after PBSC transplantation. Other strengths of our study lie in its relatively large sample size and the longterm follow-up period. We used the LT-free survival as the study end point, allowing better estimates of therapeutic efficacy of PBSC therapy. Nevertheless, several limitations were also present in this study. First, it was a retrospective study suffering from the possible selection bias. The patients who were lost to follow-up early may be different from those with complete follow-up, leading to different therapeutic effects. Thus, we performed a subgroup analysis with complete follow-up of patients and validated the performance of nomogram. Second, although the nomogram demonstrated good predictive performance, external validation was not performed. Despite advances in stem cell therapy for cirrhosis, there are still many parameters that need to be standardized from a clinical perspective. To some extent, it is challenging to find a representative cohort with a large sample size. Thus, an internal validation method was applied in our study. We plan to collect data from prospective multicenter studies to further validate our findings. Finally, we used some easily obtained clinical features



Fig. 4 Subgroup analysis of LT-free survival in patients stratified by A ALBI grade, B MELD score and C CTP class. CTP Child–Turcotte–Pugh; MELD model for end-stage liver disease, ALBI albumin–bilirubin, LT-free survival Liver-transplantation free survival

and laboratory indicators to construct this model, aiming at identifying the optimal population for PBSC therapy. However, some important parameters were not fully explored in this study. In a recently published review [35], Yang et al. placed the factors influencing the efficacy of stem cell therapy into two main categories, which were cells factors (the sources, cell preparation procedure, dosages, engraftment routes) and patients factors (liver function, inflammation, age, etc.). We could notice that there are many factors affecting the therapeutic effect. In



**Fig. 5** Influence of PBSC transplantation on the MELD score in patients listed for LT (MELD score > 15). **A** Improvement of MELD scores before and after the treatment in two groups. **B** The percentage-staked bar plots for the distribution of MELD score in two groups after PBSC therapy. *MELD* model for end-stage liver disease, *PBSC* peripheral blood stem cell, *LT* liver transplantation

the current study, on the basis of our previous research, we paid more attention to the influence of patient factors on the therapeutic effects. The cells factors remain to be considered in future extensions of this model. Furthermore, finding some valuable biomarkers from specimens of patients using transcriptomics, proteomics, and metabolomics are essential next steps in future research.

## Conclusions

In summary, based on the large-scale data derived from real-world medical records, we developed and verified a prognostic nomogram, showing good performance with individual prediction. More importantly, this risk stratification tool will be helpful to assess outcomes in clinical practice and enhance patient stratification in future clinical trials to target the optimal population.

#### Abbreviations

| PBSC             | Peripheral blood stem cell          |
|------------------|-------------------------------------|
| NLR              | Neutrophil-to-lymphocyte ratio      |
| AIC              | Akaike information criterion        |
| HBV              | Hepatitis B virus                   |
| PLT              | Platelet                            |
| ALT              | Alanine aminotransferase            |
| AST              | Aspartate aminotransferase          |
| TBIL             | Total bilirubin                     |
| PT               | Prothrombin time                    |
| INR              | International normalized ratio      |
| IQR              | Interquartile range                 |
| CI               | Confidence interval                 |
| LT-free survival | Liver-transplantation free survival |
| AUC              | Area under the curve                |
| ROC              | Receiver operator characteristic    |

## **Supplementary Information**

The online version contains supplementary material available at https://doi. org/10.1186/s13287-023-03622-y.

Additional file 1. Supplementary tables and figures.

#### Acknowledgements

Not Applicable

#### Author contributions

SYT, GYG, and XZ analyzed and interpreted the data, drafted the manuscript. GJ, KMW, and CMY contributed to the data collection. YSL, MZ, KSS, and SYM participated in the statistical analysis. LHZ, LNC, and XMZ provided academic advice. YH, YLS, and CCG were responsible for the concept and design, manuscript edit. All authors read and approved the current version of the manuscript.

#### Funding

This study was funded by National Key Research and Development Program of China (No. 2020YFA0710803 and 2017YFA0105704), National Natural Science Foundation of China (No. 81820108005, 82200680, 82270551, and 81900502) and Key Research and Development Program of Shaanxi province, China (No. 2021ZDLSF02-07 and 2022ZDLSF03-03).

#### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Declarations

#### Ethics approval and consent to participate

Informed consent was obtained from all patients and the study was approved by the Ethics Committee of Xijing Hospital on July 17, 2005. The ethics code is 20050717-1. The project title is "Autologous stem cell transplantation for the treatment of end-stage liver disease." This study conformed to the Declaration of Helsinki. This research was registered in the Chinese Clinical Trial Registry (ChiCTR-ONRC-10002070).

#### **Consent for publication**

Not applicable.

#### Competing interests

The authors declare that they have no conflicts of interests.

#### Author details

<sup>1</sup>State Key Laboratory of Cancer Biology, National Clinical Research Center for Digestive Diseases, Xijing Hospital of Digestive Diseases, Air Force Military Medical University, The Fourth Military Medical University, Xi'an 710032, Shaanxi, China. <sup>2</sup>Department of Gastroenterology, The Air Force Hospital From Eastern Theater of PLA, Nanjing 210002, Jiangsu, China.

## Received: 31 January 2023 Accepted: 20 December 2023 Published online: 02 January 2024

#### References

- Harikrishnan S, Jeemon P, Mini G, Thankappan K, Sylaja P: GBD 2017 causes of death collaborators. Global, regional, and national age-sexspecific mortality for 282 causes of death in 195 countries and territories, 1980–2017: a systematic analysis for the Global Burden of Disease Study 2017. Lancet 2018.
- D'Amico G, Morabito A, Mario P, Linda M, Giuseppe R. Clinical states of cirrhosis and competing risks. J Hepatol: J Eur Assoc Study Liver. 2018;68:563–76.
- Martin P, Dimartini A, Feng S, Brown RJ, Fallon M. Evaluation for liver transplantation in adults: 2013 practice guideline by the AASLD and the American society of transplantation. Hepatology. 2013;59(3):1144–65.
- Lucey MR. Liver transplantation in patients with alcoholic liver disease. Liver Transpl. 2011;17:751–9.
- Dwyer B, Macmillan M, Brennan P, Forbes S. Cell therapy for advanced liver diseases: repair or rebuild. J Hepatol. 2021;74:185–99.
- Alfaifi M, Eom Y, Newsome P, Baik S. Mesenchymal stromal cell therapy for liver diseases. J Hepatol. 2018;68:1272–85.
- Weissman I. Stem cells: units of development, units of regeneration, and units in evolution. Cell. 2000;100:157–68.
- am Esch J, Knoefel W, Klein M, Ghodsizad A, Fuerst G, Poll L, Piechaczek C, Burchardt E, Feifel N, Stoldt V, et al. Portal application of autologous CD133<sup>+</sup> bone marrow cells to the liver: a novel concept to support hepatic regeneration. Stem Cells (Dayton, Ohio). 2005;23:463–70.
- Lorenzini S, Isidori A, Catani L, Gramenzi A, Talarico S, Bonifazi F, Giudice V, Conte R, Baccarani M, Bernardi M, et al. Stem cell mobilization and collection in patients with liver cirrhosis. Aliment Pharmacol Ther. 2008;27:932–9.
- Pai M, Zacharoulis D, Milicevic M, Helmy S, Jiao L, Levicar N, Tait P, Scott M, Marley S, Jestice K, et al. Autologous infusion of expanded mobilized adult bone marrow-derived CD34<sup>+</sup> cells into patients with alcoholic liver cirrhosis. Am J Gastroenterol. 2008;103:1952–8.
- Han Y, Yan L, Han G, Zhou X, Hong L, Yin Z, Zhang X, Wang S, Wang J, Sun A, et al. Controlled trials in hepatitis B virus-related decompensate liver cirrhosis: peripheral blood monocyte transplant versus granulocyte-colony-stimulating factor mobilization therapy. Cytotherapy. 2008;10:390–6.
- Salama H, Zekri A, Zern M, Bahnassy A, Loutfy S, Shalaby S, Vigen C, Burke W, Mostafa M, Medhat E, et al. Autologous hematopoietic stem cell transplantation in 48 patients with end-stage chronic liver diseases. Cell Transplant. 2010;19:1475–86.
- Levicar N, Pai M, Habib N, Tait P, Jiao L, Marley S, Davis J, Dazzi F, Smadja C, Jensen S, et al. Long-term clinical results of autologous infusion of mobilized adult bone marrow derived CD34<sup>+</sup> cells in patients with chronic liver disease. Cell Prolif. 2008;41:115–25.
- Newsome P, Fox R, King A, Barton D, Than N, Moore J, Corbett C, Townsend S, Thomas J, Guo K, et al. Granulocyte colony-stimulating factor and autologous CD133-positive stem-cell therapy in liver cirrhosis (REALISTIC): an open-label, randomised, controlled phase 2 trial. Lancet Gastroenterol Hepatol. 2018;3:25–36.

- Guo C, Guo G, Zhou X, Chen Y, Han Z, Yang C, Zhao S, Su H, Lian Z, Leung P, et al. Long-term outcomes of autologous peripheral blood stem cell transplantation in patients with cirrhosis. Clin Gastroenterol Hepatol: Off Clin Pract J Amer Gastroenterol Assoc. 2019;17:1175-1182. e1172.
- Angeli P, Bernardi M, Villanueva C, Francoz C, Mookerjee RP, Trebicka J, Krag A, Laleman W, Gines P. EASL clinical practice guidelines for the management of patients with decompensated cirrhosis. J Hepatol. 2018;69:406–60.
- Qureshi W, Adler D, Davila R, Egan J, Hirota W, Leighton J, Rajan E, Zuckerman M, Fanelli R, Wheeler-Harbaugh J, et al. ASGE guideline: the role of endoscopy in the management of variceal hemorrhage, updated July 2005. Gastrointest Endosc. 2005;62:651–5.
- Biggins S, Angeli P, Garcia-Tsao G, Ginès P, Ling S, Nadim M, Wong F, Kim W. Diagnosis, evaluation, and management of ascites, spontaneous bacterial peritonitis and hepatorenal syndrome: 2021 practice guidance by the american association for the study of liver diseases. Hepatology (Baltimore, MD). 2021;74:1014–48.
- Cholongitas E, Papatheodoridis GV, Vangeli M, Terreni N, Patch D, Burroughs AK. Systematic review: the model for end: stage liver disease: should it replace Child-Pugh's classification for assessing prognosis in cirrhosis? Aliment Pharmacol Ther. 2005;22:1079–89.
- Johnson P, Berhane S, Kagebayashi C, Satomura S, Teng M, Reeves H, O'Beirne J, Fox R, Skowronska A, Palmer D, et al. Assessment of liver function in patients with hepatocellular carcinoma: a new evidencebased approach-the ALBI grade. J Clin Oncol: Off J Am Soc Clin Oncol. 2015;33:550–8.
- 21. Kamath PS, Kim WR. Advanced liver disease study group. The model for end-stage liver disease (MELD). Hepatology. 2007;45:797–805.
- 22. Lee CW, Chen YF, Wu HH, Lee OK. Historical perspectives and advances in mesenchymal stem cell research for the treatment of liver diseases. Gastroenterology. 2018;154(1):46–56.
- Dwyer BJ, Macmillan MT, Brennan PN, Forbes SJ. Cell therapy for advanced liver diseases: repair or rebuild—ScienceDirect. J Hepatol. 2021;74:185–99.
- Shi M, Li Y, Xu R, Meng F, Yu S, Fu J, Hu J, Li J, Wang L, Jin L, Wang F. Mesenchymal stem cell therapy in decompensated liver cirrhosis: a long-term follow-up analysis of the randomized controlled clinical trial. Hepatol Int. 2021;15:1431–41.
- Zhang K, Sun H, Cao H, Jia Y, Shu X, Cao H, Zhang Y, Yang X. The impact of recipient age on the effects of umbilical cord mesenchymal stem cells on HBV-related acute-on-chronic liver failure and liver cirrhosis. Stem Cell Res Ther. 2021;12:466.
- Biyik M, Ucar R, Solak Y, Gungor G, Polat I, Gaipov A, Cakir O, Ataseven H, Demir A, Turk S, Polat H. Blood neutrophil-to-lymphocyte ratio independently predicts survival in patients with liver cirrhosis. Eur J Gastroenterol Hepatol. 2013;25:435–41.
- Leithead J, Rajoriya N, Gunson B, Ferguson J. Neutrophil-to-lymphocyte ratio predicts mortality in patients listed for liver transplantation. Liver Int: Off J Int Assoc Study Liver. 2015;35:502–9.
- Honggang Z, Qinqin S, Weilin M, Jian F, Bo Y. Neutrophil-to-lymphocyte ratio predicts early mortality in patients with HBV-related decompensated cirrhosis. Gastroenterol Res Pract. 2016;2016:1–5.
- Shi Y, Wang Y, Li Q, Liu K, Hou J, Shao C, Wang Y. Immunoregulatory mechanisms of mesenchymal stem and stromal cells in inflammatory diseases. Nat Rev Nephrol. 2018;14:493–507.
- Verma N, Singh A, Singh V. Haematopoietic stem cells in cirrhosis. Lancet Gastroenterol Hepatol. 2018;3:298.
- Thuluvath P, Guidinger M, Fung J, Johnson L, Rayhill S, Pelletier S. Liver transplantation in the United States, 1999–2008. Am J Transplant Off J Am Soc Transplant Am Soc Transplant Surg. 2010;10:1003–19.
- Kim W, Therneau T, Benson J, Kremers W, Rosen C, Gores G, Dickson E. Deaths on the liver transplant waiting list: an analysis of competing risks. Hepatology (Baltimore, MD). 2006;43:345–51.
- 33. Freeman R, Wiesner R, Harper A, McDiarmid S, Lake J, Edwards E, Merion R, Wolfe R, Turcotte J, Teperman L. The new liver allocation system: moving toward evidence-based transplantation policy. Liver Transplant: Off Publ Am Assoc Study Liver Dis Int Liver Transplant Soc. 2002;8:851–8.
- Martin P, DiMartini A, Feng S, Brown R, Fallon M. Evaluation for liver transplantation in adults: 2013 practice guideline by the American

association for the study of liver diseases and the american society of transplantation. Hepatology (Baltimore, MD). 2014;59:1144–65.

 Yang X, Li Q, Liu W, Zong C, Wei L, Shi Y, Han Z. Mesenchymal stromal cells in hepatic fibrosis/cirrhosis: from pathogenesis to treatment. Cell Mol Immunol. 2023;20:583–99.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

## Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

#### At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

